

1 **Performance Characteristics of BinaxNOW COVID-19 Antigen Card for Screening**

2 **Asymptomatic Individuals in a University Setting**

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21 for the study together with the senior author.

22

23 ABSTRACT

24 We compared the performance of the Abbott BinaxNOW COVID-19 Antigen Card to a standard
25 RT-PCR assay (ThermoFisher TaqPath COVID-19 Combo Kit) for the detection of SARS-CoV-2 in
26 2,645 asymptomatic students presenting for screening at the University of Utah. SARS-CoV-2
27 RNA was detected in 1.7% of the study participants by RT-PCR. BinaxNOW identified 24
28 infections but missed 21 infections that were detected by RT-PCR. The analytical sensitivity
29 (positive agreement) and analytical specificity (negative agreement) for the BinaxNOW was
30 53.3% and 100%, respectively when compared against the RT-PCR assay. The median cycle
31 threshold (Ct) value in the specimens that had concordant positive BinaxNOW antigen result
32 was significantly lower compared to those that were discordant (Ct 17.6 vs. 29.6; $p < 0.001$). In
33 individuals with presumably high viral loads (Ct < 23.0), a 95.8% positive agreement was
34 observed between the RT-PCR assay and BinaxNOW. Due to the possibility of false negative
35 results, caution must be taken when utilizing rapid antigen testing for screening asymptomatic
36 individuals.

37

38 INTRODUCTION

39 With its high degree of transmissibility, the Severe Acute Respiratory Syndrome
40 Coronavirus 2 (SARS-CoV-2), the causative pathogen for the novel 2019 coronavirus disease
41 (COVID-19), has undoubtedly led to one of the most remarkable global public health epidemics
42 in recent history. Timely identification and isolation of infected individuals is crucial in
43 mitigating rampant community spread of SARS-CoV-2. The gold standard method for COVID-19
44 diagnosis remains detection of SARS-CoV-2 ribonucleic acid (RNA) in respiratory tract specimens

45 using nucleic acid amplification techniques such as reverse transcription polymerase chain
46 reaction (RT-PCR). However, SARS-CoV-2 nucleic acid amplification tests (NAAT) are generally
47 more expensive than alternative methodologies and may have prolonged turnaround times due
48 to limited test supplies, reagent allocation, and fixed laboratory capacity, which have been
49 exacerbated by extremely high demand.

50 Efforts to expand testing capacity have led to the development of several rapid antigen
51 tests designed to detect SARS-CoV-2 nucleocapsid antigen, primarily in symptomatic individuals
52 (1). At the time of this writing, the United States Food and Drug Administration (FDA) has
53 granted emergency use authorization (EUA) to eleven SARS-CoV-2 antigen tests (2). Although
54 these antigen tests are intended to be utilized in symptomatic individuals (within the first five
55 to seven days of symptom onset), the United States Department of Health and Human Services
56 (HHS), through the Public Readiness and Emergency Preparedness Act (PREP Act), permits their
57 use for screening asymptomatic individuals in congregate facilities, including schools (3).
58 However, there is limited data on the performance characteristics of rapid antigen tests in
59 asymptomatic or pre-symptomatic individuals. A recent meta-analysis of published literature on
60 rapid, point-of-care antigen tests reported an average sensitivity and specificity of 56.2% and
61 99.5%, respectively, when compared to NAAT (1). However, these studies were not limited
62 exclusively to asymptomatic individuals, the specimen type was primarily nasopharyngeal
63 and/or oropharyngeal, and none of the antigen tests included have received EUA approval from
64 the FDA.

65 In this study, we evaluated the diagnostic performance characteristics of the Abbott
66 BinaxNOW COVID-19 Antigen Card (hereby referred to as BinaxNOW) in a population of college-

67 age students who were asymptomatic at the time of testing. BinaxNOW is a rapid lateral flow
68 immunoassay that qualitatively detects SARS-CoV-2 nucleocapsid antigen in direct nasal swab
69 specimens. The package insert cites a positive agreement of 97.1% and a negative agreement of
70 98.5% when compared against an EUA RT-PCR assay (4). These data were based on a clinical
71 study involving a total of 102 patients, of which 95 had symptoms consistent with COVID-19
72 and only 7 were asymptomatic. This was recently updated to a positive agreement of 84.6%,
73 based on a larger study involving 460 symptomatic individuals. Of note, the United States
74 federal government has distributed 150 million BinaxNOW Antigen Cards to states across the
75 country (5). BinaxNOW also received EUA for at-home use under the supervision of a telehealth
76 proctor (6). Therefore, characterizing the performance characteristics of BinaxNOW for off-label
77 use in an asymptomatic population is essential given its potential widespread application for
78 asymptomatic screening in a variety of settings.

79

80 MATERIALS AND METHODS

81 **Study population and specimen collection.**

82 The participants of this study were primarily college-age (undergraduate and graduate)
83 students at the University of Utah in Salt Lake City, Utah, USA. At the time of specimen
84 collection, the students were first queried to ensure that they were not experiencing any signs
85 and/or symptoms of COVID-19. Specimen collection occurred at a temporary indoor testing site
86 from November 13-20, 2020. Two nasal swabs were collected from each participant, following
87 the technique recommended by the United States Center for Disease Control and Prevention
88 (CDC) (7). The study participants were instructed to swab both nares at the level of the mid

89 turbinate for each collection. Trained non-medical personnel observed the specimen collection
90 process. The first swab collected from the participants was randomly assigned to be tested
91 either with BinaxNOW or the RT-PCR assay in an effort to minimize sampling bias.

92 **Detection of SARS-CoV-2 viral antigen**

93 The BinaxNOW Antigen Cards utilized in this study were received from the Utah
94 Department of Health as part of a United States federal government initiative to expand COVID-
95 19 testing capacity. Testing was performed by trained non-medical personnel (University of
96 Utah Hope Corps Interns) according to the manufacturer's instructions (4). Each testing
97 personnel was trained on the test procedure (including appropriate use of personal protective
98 equipment) and result interpretation using detailed step-by-step videos provided by the
99 manufacturer. To evaluate for competence, each testing personnel was required to pass an
100 assessment quiz and successfully perform external quality control using a positive control swab
101 and a sterile swab (negative control). External quality control was also performed for each new
102 kit of BinaxNOW Antigen Cards.

103 Results were interpreted visually after 15 minutes. A specimen was deemed positive for
104 SARS-CoV-2 viral antigen if two pink/purple colored lines (control line on the top and sample
105 line on the bottom) were observed on the test card, as illustrated in the assay product insert
106 (4). A faint pink/purple colored line in the sample region of the test card (in addition to a
107 pink/purple colored control line) was also interpreted as a positive result. A single pink/purple
108 colored line in the control region of the test card was interpreted as a negative result. If no line
109 was observed in the control region or if the line remains blue in color, then the result was
110 interpreted as invalid.

111 Participants were notified of their BinaxNOW result using the NAVICA Mobile App,
112 which is a free mobile app provided by Abbott (8). Any participant that tested positive was
113 contacted to return to the testing site within 24 hours and submit a saliva specimen for SARS-
114 CoV-2 NAAT at ARUP Laboratories. These individuals were instructed to self-isolate while
115 awaiting NAAT confirmation. Individuals that received an invalid BinaxNOW result were also
116 contacted for repeat antigen testing. Participants receiving a negative antigen test were
117 counseled that these results were “presumptive” and did not negate the need for mitigation
118 behaviors designed to reduce the spread of SARS-CoV-2.

119 **Detection of SARS-CoV-2 nucleic acid**

120 The other nasal swab was placed into ARUP COVID-19 Transport Media™ (9) and tested
121 at ARUP Laboratories using the ThermoFisher TaqPath COVID-19 Combo Kit, hereby referred to
122 as the TaqPath COVID-19 Kit (10). These specimens were stored frozen (-20 °C) and tested
123 within 10 days of receipt in the clinical laboratory. The TaqPath COVID-19 Kit targets regions of
124 three coronavirus genes: ORF1ab, the gene for the S protein, and the gene for the N protein. 40
125 amplification cycles are performed by the assay. At least two genes have to be detected for the
126 result to be reported as positive for SARS-CoV-2. The cycle threshold (Ct) value for each
127 specimen was reported as the average of the Ct values of the detected coronavirus genes. An
128 inconclusive result was reported when only one gene is detected after consecutive repeat
129 testing. Detection of SARS-CoV-2 RNA in the confirmatory saliva specimens was performed in
130 real-time using one of three FDA EUA assays (either Hologic Panther Fusion SARS-CoV-2 assay,
131 Roche Cobas SARS-CoV-2 assay, or ThermoFisher TaqPath COVID-19 Combo Kit). All participants
132 were notified of their NAAT results.

133

134 **Statistical analysis**

135 The TaqPath COVID-19 Kit was used as the benchmark for assessing the diagnostic
136 accuracy of BinaxNOW. The analytical performance characteristics (sensitivity, specificity, and
137 predictive values) were calculated from a 2x2 contingency table using GraphPad Prism 8
138 software. Agreement between methods was assessed at various Ct cutoffs reported in the
139 package insert for BinaxNOW (4) and published literature. The 95% confidence intervals are
140 based on the Wilson-Brown method. A non-parametric t test (Mann-Whitney test) was
141 performed using GraphPad Prism 8 software to evaluate for statistical significance (p values)
142 between median Ct values. Kappa coefficient was calculated using the Microsoft Excel Analyse-
143 it software package (version 5.20).

144

145 **RESULTS**146 **Positivity rate of the rapid antigen test and nucleic acid amplification test**

147 Two nasal swab specimens were collected from 2,645 individuals. Among the study
148 participants, 1369 (51.8%) identified as female, 1274 (48.2%) identified as male, while 2 (0.1%)
149 identified as non-binary. The average age of the study participants was 24 years (range: 15 to
150 86 years). **Table 1** summarizes the results from BinaxNOW and the TaqPath COVID-19 Kit. A
151 negative result with BinaxNOW was observed in 2,618 (99.0%) individuals, while a positive
152 result was observed in 24 (0.9%) individuals. An invalid BinaxNOW result was initially observed
153 in 3 (0.1%) individuals; however, repeat testing using a new nasal swab specimen from these
154 individuals yielded a negative result. For the TaqPath COVID-19 Kit, SARS-CoV-2 RNA was not

155 detected in 2,595 (98.1%) individuals, 46 (1.7%) individuals had detectable SARS-CoV-2 RNA,
156 while 4 (0.2%) individuals had an inconclusive result.

157 **Concordance between the rapid antigen test and the nucleic acid amplification test**

158 The analytical sensitivity and specificity of BinaxNOW is summarized in **Table 2**. Of the
159 46 individuals that had detectable SARS-CoV-2 RNA, 24 had a concordant positive antigen
160 result, indicating a positive agreement of 53.3% between the two tests. The kappa coefficient (κ
161 0.69; 95% CI: 0.57 – 0.82) indicates substantial agreement between methods. The median cycle
162 threshold (Ct) value in the specimens that had concordant positive results was significantly
163 lower (Ct 17.6) than those that were discordant (Ct 29.6) ($p < 0.001$), as illustrated in **Figure 1**.
164 In specimens with presumably high viral loads (Ct < 23.0), a 95.8% positive agreement was
165 observed (**Table 3**). A 0% positive agreement was observed in samples with both Ct ≥ 33 and Ct
166 ≥ 30 , as shown in **Table 3**.

167 Collection of two consecutive bilateral nasal swab specimens did not significantly affect
168 the detection of SARS-CoV-2 using either NAAT or the rapid antigen test ($p = 0.5683$; Fisher's
169 exact test). The rapid antigen test was performed using the first nasal swab specimen in 12
170 (50%) out of the 24 individuals with concordant positive results. No statistically significant
171 difference in median Ct value was observed in concordant positive samples regardless of
172 whether the rapid antigen test was performed using the first nasal swab versus the second
173 nasal swab (**Figure 2**) ($p = 0.5800$). A discordant result between the rapid antigen test and
174 NAAT (i.e., antigen negative/NAAT positive) was observed in 21 individuals. Discordant results
175 between BinaxNOW and the RT-PCR assay were more likely at Ct values > 23.0 , as shown **Figure**
176 **3**. The antigen test was performed using the first nasal swab specimen in 9 (40.9%) out of the

177 21 individuals with discordant results. While a slightly higher median Ct value was observed
178 when the antigen test was performed using the second nasal swab versus the first nasal swab,
179 the difference was not statistically significant ($p = 0.1752$), as shown in **Figure 2**. In one
180 individual with a discordant result, an invalid BinaxNOW antigen result was initially obtained,
181 with a negative result observed upon repeat testing using a new nasal swab specimen. It is
182 worth mentioning that for this individual, the initial invalid BinaxNOW was obtained using the
183 second nasal swab specimen, while the negative result from the repeat test was obtained from
184 a third nasal swab. Hence, the validity of the negative BinaxNOW result in this individual could
185 be questionable due to sampling bias. Invalid results were excluded in the diagnostic
186 performance characteristics calculations.

187 Twenty-two out of the 24 individuals (91.7%) with a positive antigen result returned to
188 the testing site and submitted a follow-up saliva specimen. There was 100% agreement
189 between these positive BinaxNOW specimens and saliva NAAT.

190

191 DISCUSSION

192 When compared to NAAT, the BinaxNow Antigen Card showed low analytical sensitivity
193 (53.3%) for detecting SARS-CoV-2 infection in an asymptomatic or pre-symptomatic population.
194 This observation is consistent with the findings of other recent studies conducted using
195 different SARS-CoV-2 antigen assays in unselected populations (11-13). Collection of two
196 consecutive bilateral nasal swab specimens did not statistically affect the detection of SARS-
197 CoV-2 using either the RT-PCR assay or the rapid antigen test. However, there was a trend
198 toward higher Ct values in the second swab indicating a lesser amount of virus present, which

199 may have disproportionately affected the antigen positivity rate. One study found a difference
200 of 6-7 Ct between the limit of detection of the BinaxNOW antigen test and RT-PCR tests,
201 indicating an approximate 100-fold difference in sensitivity (14).

202 Our results indicate that a relatively high viral load (and corresponding low Ct value <23)
203 must be present to generate a positive BinaxNOW result. At the onset of our study, the
204 BinaxNOW product insert reported a positive agreement of 83.3% in specimens with Ct \geq 33 (4).
205 The manufacturer has recently updated this information to a positive agreement of 37.8%. Ct
206 values are a relative approximation of virus load. Differences in assay design and other
207 important pre-analytic variables (e.g., specimen source, collection method, volume of transport
208 media, etc.) impact reported Ct values such that these measurements are not directly
209 comparable across real-time NAAT platforms (15).

210 In contrast to analytical sensitivity, the specificity of BinaxNOW testing was excellent
211 (100%). The test was able to be performed successfully at the point of care by non-medical
212 personnel with a relatively low invalid rate (0.1%), supporting the findings of another recently
213 published study (16). These observations raise the question of whether confirmation of positive
214 BinaxNOW results is necessary, as cautioned in a recent warning by the FDA regarding the
215 potential for false positive results from rapid SARS-CoV-2 antigen tests (17). It is important to
216 note, however, that operators underwent comprehensive training and quality control testing
217 was performed regularly on-site. This is especially important in the context of at home testing.
218 Additional studies are needed to determine whether BinaxNOW test performance will be
219 comparable in a telehealth-observed home setting.

220 Despite its relatively low analytical sensitivity, BinaxNOW may still be beneficial for
221 surveillance testing in selected settings where testing resources are limited, especially when
222 weighed against the alternative of no screening testing. Rapid antigen testing identified 24
223 infections in asymptomatic individuals, with qualitatively high viral loads, who may be more
224 likely to be infectious to others (18, 19). These infections were all confirmed by saliva NAAT
225 and individuals were instructed to self-isolate. Given the relatively low prevalence (1.7%) in our
226 student population, the negative predictive value of BinaxNow was excellent (99.2%).

227 A total of 21 asymptomatic students had false negative antigen tests. We do not know if
228 these individuals developed symptoms in the days following the negative antigen result. We
229 also cannot speculate as to how infectious these individuals were; presumably, the risk of viral
230 transmission to others is not zero (18, 19) although the higher Ct values associated with these
231 samples may indicate a low risk of transmission. However, it is well established that
232 asymptomatic carriers of SARS-CoV-2 can efficiently transmit the infection (20, 21). Thus, all
233 participants were counseled to continue with physical distancing, face masking, and proper
234 hand hygiene despite a negative BinaxNOW result. The public health implications of a false
235 negative screening result in an asymptomatic population will depend on the population to
236 which the test is applied. For example, tolerance for false negatives may be greater in a
237 congregate setting consisting of young, otherwise healthy individuals (e.g., college campus)
238 with few risk factors for severe clinical outcome from COVID-19 versus a long-term care facility
239 setting or other demographic with one or multiple risk factors for poor COVID-19 associated
240 outcomes.

241 The limitations of this study include the relatively small number of positive results and lack
242 of serial repeat testing data for the asymptomatic student cohort to determine if the 21 false
243 negatives result would eventually test positive after subsequent assessments. This would be
244 useful for validating the effectiveness of the proposed strategy of repeat serial testing using less
245 sensitive antigen tests as an infection prevention and control measure (22, 23).

246 To the best of our knowledge, this is the first study evaluating the performance of a rapid
247 SARS-CoV-2 antigen test in an exclusively asymptomatic population. The analytical sensitivity of
248 BinaxNOW for off-label use in an asymptomatic population is lower than the performance
249 claims for symptomatic patients reported by the manufacturer. As recommended by the
250 manufacturer, negative results should be interpreted as presumptive negative. Careful
251 assessment of the impact of false negative results is warranted before a testing strategy
252 utilizing rapid SARS-CoV-2 antigen tests is implemented. The specificity BinaxNOW, however,
253 was excellent.

254

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259 student body for participating in this study.

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263 REFERENCES

- 264 1. Dinnes J, Deeks JJ, Adriano A, Berhane S, Davenport C, Dittrich S, Emperador D,
265 Takwoingi Y, Cunningham J, Beese S, et al. 2020. Rapid, point-of-care antigen and
266 molecular-based tests for diagnosis of SARS-CoV-2 infection. *Cochrane Database Syst*
267 *Rev* doi:10.1002/14651858.CD013705.
- 268 2. U.S. Food and Drug Administration. Individual EUAs for Antigen Diagnostic Tests for
269 SARS-CoV-2. [https://www.fda.gov/medical-devices/coronavirus-disease-2019-covid-19-](https://www.fda.gov/medical-devices/coronavirus-disease-2019-covid-19-emergency-use-authorizations-medical-devices/vitro-diagnostics-euas#individual-antigen)
270 [emergency-use-authorizations-medical-devices/vitro-diagnostics-euas#individual-](https://www.fda.gov/medical-devices/coronavirus-disease-2019-covid-19-emergency-use-authorizations-medical-devices/vitro-diagnostics-euas#individual-antigen)
271 [antigen](https://www.fda.gov/medical-devices/coronavirus-disease-2019-covid-19-emergency-use-authorizations-medical-devices/vitro-diagnostics-euas#individual-antigen). Accessed December 22, 2020.
- 272 3. U.S. Department of Health and Human Services. Guidance for PREP Act Coverage for
273 COVID-19 Screening Tests at Nursing Homes, Assisted-Living Facilities, Long-Term-Care
274 Facilities, and other Congregate Facilities.
275 [https://www.hhs.gov/guidance/sites/default/files/hhs-guidance-documents//prep-act-](https://www.hhs.gov/guidance/sites/default/files/hhs-guidance-documents//prep-act-coverage-for-screening-in-congregate-settings.pdf)
276 [coverage-for-screening-in-congregate-settings.pdf](https://www.hhs.gov/guidance/sites/default/files/hhs-guidance-documents//prep-act-coverage-for-screening-in-congregate-settings.pdf). Accessed December 19, 2020.
- 277 4. BinaxNOW COVID-19 Ag Card [Product Insert]. Scarborough, ME: Abbott Diagnostics
278 Scarborough, Inc; 2020.
- 279 5. U.S. Department of Health and Human Services. Trump Administration Will Deploy 150
280 Million Rapid Tests in 2020. [https://www.hhs.gov/about/news/2020/08/27/trump-](https://www.hhs.gov/about/news/2020/08/27/trump-administration-will-deploy-150-million-rapid-tests-in-2020.html)
281 [administration-will-deploy-150-million-rapid-tests-in-2020.html](https://www.hhs.gov/about/news/2020/08/27/trump-administration-will-deploy-150-million-rapid-tests-in-2020.html). Accessed December 19,
282 2020.
- 283 6. U.S. Food and Drug Administration. BinaxNOW COVID-19 Ag Card Home Test.
284 <https://www.fda.gov/media/144576/download>. Accessed December 21, 2020.
- 285 7. U.S. Center for Disease Control and Prevention. Interim Guidelines for Collecting,
286 Handling, and Testing Clinical Specimens for COVID-19.
287 <https://www.cdc.gov/coronavirus/2019-ncov/lab/guidelines-clinical-specimens.html>.
288 Accessed December 19, 2020.
- 289 8. Abbott. Your NAVICA App Questions Answered. [https://www.abbott.com/BinaxNOW-](https://www.abbott.com/BinaxNOW-Test-NAVICA-App/NAVICA-FAQ.html)
290 [Test-NAVICA-App/NAVICA-FAQ.html](https://www.abbott.com/BinaxNOW-Test-NAVICA-App/NAVICA-FAQ.html). Accessed December 23, 2020.
- 291 9. Hanson KE, Barker AP, Hillyard DR, Gilmore N, Barrett JW, Orlandi RR, Shakir SM. 2020.
292 Self-Collected Anterior Nasal and Saliva Specimens versus Health Care Worker-Collected
293 Nasopharyngeal Swabs for the Molecular Detection of SARS-CoV-2. *J Clin Microbiol*
294 58:e01824-20.
- 295 10. U.S. Food and Drug Administration. TaqPath COVID-19 Combo Kit.
296 <https://www.fda.gov/media/136113/download>. Accessed December 19, 2020.
- 297 11. Lambert-Niclot S, Cuffel A, Le Pape S, Vauloup-Fellous C, Morand-Joubert L, Roque-
298 Afonso A-M, Le Goff J, Delaugerre C. 2020. Evaluation of a Rapid Diagnostic Assay for
299 Detection of SARS-CoV-2 Antigen in Nasopharyngeal Swabs. *J Clin Microbiol* 58:e00977-
300 20.
- 301 12. Scohy A, Anantharajah A, Bodéus M, Kabamba-Mukadi B, Verroken A, Rodriguez-
302 Villalobos H. 2020. Low performance of rapid antigen detection test as frontline testing
303 for COVID-19 diagnosis. *J Clin Virol* 129:104455.

- 304 13. Mak GCK, Lau SSY, Wong KKY, Chow NLS, Lau CS, Lam ETK, Chan RCW, Tsang DNC. 2020.
305 Analytical sensitivity and clinical sensitivity of the three rapid antigen detection kits for
306 detection of SARS-CoV-2 virus. *J Clin Virol* 133:104684.
- 307 14. Perchetti GA, Huang M-L, Mills MG, Jerome KR, Greninger AL. 2020. Analytical
308 Sensitivity of the Abbott BinaxNOW COVID-19 Ag CARD. *J Clin Microbiol*
309 doi:10.1128/jcm.02880-20:JCM.02880-20.
- 310 15. Rhoads D, Peaper DR, She RC, Nolte FS, Wojewoda CM, Anderson NW, Pritt BS. 2020.
311 College of American Pathologists (CAP) Microbiology Committee Perspective: Caution
312 Must Be Used in Interpreting the Cycle Threshold (Ct) Value. *Clin Infect Dis*
313 doi:10.1093/cid/ciaa1199.
- 314 16. Pilarowski G, Lebel P, Sunshine S, Liu J, Crawford E, Marquez C, Rubio L, Chamie G,
315 Martinez J, Peng J, Black D, Wu W, Pak J, Laurie MT, Jones D, Miller S, Jacobo J, Rojas S,
316 Rojas S, Nakamura R, Tulier-Laiwa V, Petersen M, Havlir DV, DeRisi J. 2020. Performance
317 characteristics of a rapid SARS-CoV-2 antigen detection assay at a public plaza testing
318 site in San Francisco. medRxiv doi:10.1101/2020.11.02.20223891:2020.11.02.20223891.
- 319 17. U.S. Food and Drug Administration. Potential for False Positive Results with Antigen
320 Tests for Rapid Detection of SARS-CoV-2 - Letter to Clinical Laboratory Staff and Health
321 Care Providers. [https://www.fda.gov/medical-devices/letters-health-care-
322 providers/potential-false-positive-results-antigen-tests-rapid-detection-sars-cov-2-
323 letter-clinical-laboratory](https://www.fda.gov/medical-devices/letters-health-care-providers/potential-false-positive-results-antigen-tests-rapid-detection-sars-cov-2-letter-clinical-laboratory). Accessed December 19, 2020.
- 324 18. Bullard J, Dust K, Funk D, Strong JE, Alexander D, Garnett L, Boodman C, Bello A, Hedley
325 A, Schiffman Z, Doan K, Bastien N, Li Y, Van Caesele PG, Poliquin G. 2020. Predicting
326 Infectious Severe Acute Respiratory Syndrome Coronavirus 2 From Diagnostic Samples.
327 *Clin Infect Dis* 71:2663-2666.
- 328 19. Basile K, McPhie K, Carter I, Alderson S, Rahman H, Donovan L, Kumar S, Tran T, Ko D,
329 Sivaruban T, Ngo C, Toi C, O'Sullivan MV, Sintchenko V, Chen SC-A, Maddocks S, Dwyer
330 DE, Kok J. 2020. Cell-based culture of SARS-CoV-2 informs infectivity and safe de-
331 isolation assessments during COVID-19. *Clin Infect Dis* doi:10.1093/cid/ciaa1579.
- 332 20. Sugano N, Ando W, Fukushima W. 2020. Cluster of Severe Acute Respiratory Syndrome
333 Coronavirus 2 Infections Linked to Music Clubs in Osaka, Japan. *J Infect Dis* 222:1635-
334 1640.
- 335 21. Arons MM, Hatfield KM, Reddy SC, Kimball A, James A, Jacobs JR, Taylor J, Spicer K,
336 Bardossy AC, Oakley LP, Tanwar S, Dyal JW, Harney J, Chisty Z, Bell JM, Methner M, Paul
337 P, Carlson CM, McLaughlin HP, Thornburg N, Tong S, Tamin A, Tao Y, Uehara A, Harcourt
338 J, Clark S, Brostrom-Smith C, Page LC, Kay M, Lewis J, Montgomery P, Stone ND, Clark
339 TA, Honein MA, Duchin JS, Jernigan JA. 2020. Presymptomatic SARS-CoV-2 Infections
340 and Transmission in a Skilled Nursing Facility. *N Engl J Med* 382:2081-2090.
- 341 22. Mina MJ, Parker R, Larremore DB. 2020. Rethinking Covid-19 Test Sensitivity — A
342 Strategy for Containment. *N Engl J Med* 383:e120.
- 343 23. U.S. Center for Disease Control and Prevention. Considerations for Use of SARS-CoV-2
344 Antigen Testing in Nursing Homes. [https://www.cdc.gov/coronavirus/2019-
345 ncov/hcp/nursing-homes-antigen-testing.html](https://www.cdc.gov/coronavirus/2019-ncov/hcp/nursing-homes-antigen-testing.html). Accessed December 22, 2020.
- 346

347 **Figure Legends**

348 Figure 1: Distribution of the RT-PCR cycle threshold (Ct) values in specimens with positive and
349 negative BinaxNOW results. p value is based on the Mann-Whitney test. The lines signify
350 median and interquartile ranges.

351 Figure 2: Distribution of the RT-PCR cycle threshold (Ct) values in specimens with concordant
352 positive BinaxNOW results (**A**) and discordant negative BinaxNOW results (**B**) sorted by order of
353 nasal swab collection. p value is based on the Mann-Whitney test. The lines signify median and
354 interquartile ranges.

355 Figure 3: Frequency distribution of RT-PCR cycle threshold (Ct) values in all specimens with
356 detectable SARS-CoV-2 and specimens with discordant BinaxNOW results.

Table 1. Summary of results from the BinaxNOW Antigen Card and the TaqPath COVID-19

Kit

	BinaxNOW Antigen Card	TaqPath COVID-19 Kit
Positive	24	46
Negative	2618	2595
Inconclusive / Invalid	3*	4 [#]
Total	2645	2645
*Repeat testing yielded a negative result		
[#] Only the N protein gene was detected in these specimens (Ct value was > 30)		

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369 **Table 2.** Diagnostic performance of BinaxNOW Antigen Card compared to TaqPath COVID-19 Kit

370 for detection of SARS-CoV-2

BinaxNOW Antigen Card	TaqPath COVID-19 Kit		
	Positive	Negative	Total
Positive	24	0	24
Negative	21	2593	2614
Total	45	2593	2638
Analytical sensitivity (positive agreement) = 53.3% (95% CI: 39.1% – 67.1%)			
Analytical specificity (negative agreement) = 100% (95% CI: 99.9% – 100%)			
Positive predictive value* = 100% (95% CI: 86.2% – 100%)			
Negative predictive value* = 99.2% (95% CI: 98.7% – 99.4%)			
Kappa coefficient = 0.69 (95% CI: 0.57 – 0.82)			
*Predictive values are assuming a disease prevalence of 1.7%			
Note: 4 inconclusive RT-PCR results and 3 invalid BinaxNOW results were excluded from the calculations above			

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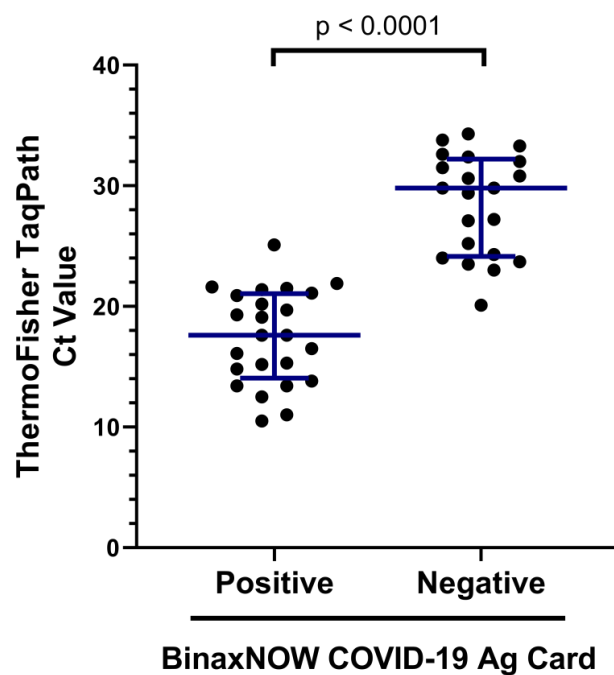
377 **Table 3.** BinaxNOW Antigen Card diagnostic performance against the comparator RT-PCR method

378 by cycle threshold counts

BinaxNOW Antigen Card	TaqPath COVID-19 Kit (Positive Results by Ct Category)					
	Ct < 33.0	Ct ≥ 33.0	Ct < 30.0	Ct ≥ 30.0	Ct < 23.0	Ct ≥ 23.0
Positive	24	0	24	0	23	1
Negative	18	3	12	9	1	20
Total	42	3	36	9	24	21
Positive Agreement (95% CI)	57.1% (42.2 – 70.9)	0%	66.7% (50.3 – 79.8)	0%	95.8% (79.8 – 99.3)	4.8% (0.8 – 22.7)

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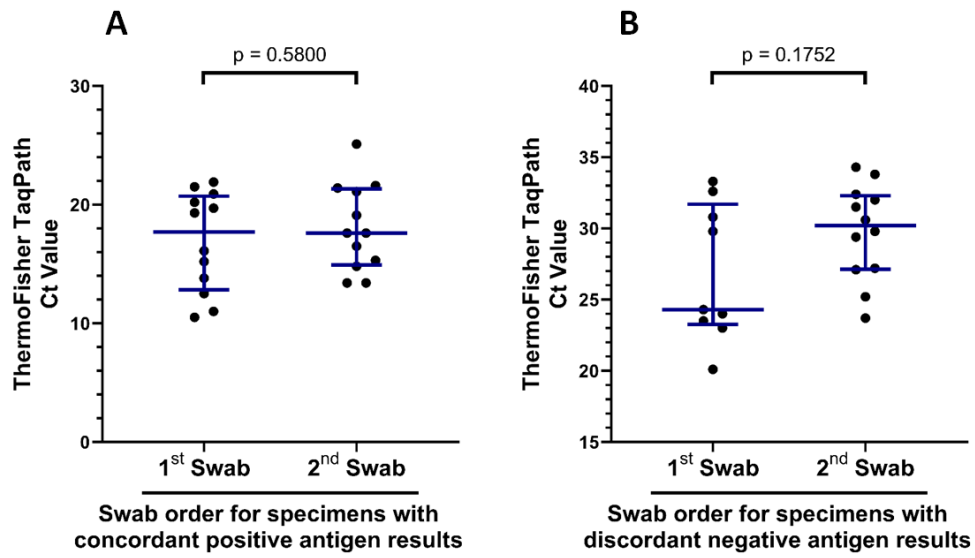
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382 Figure 1

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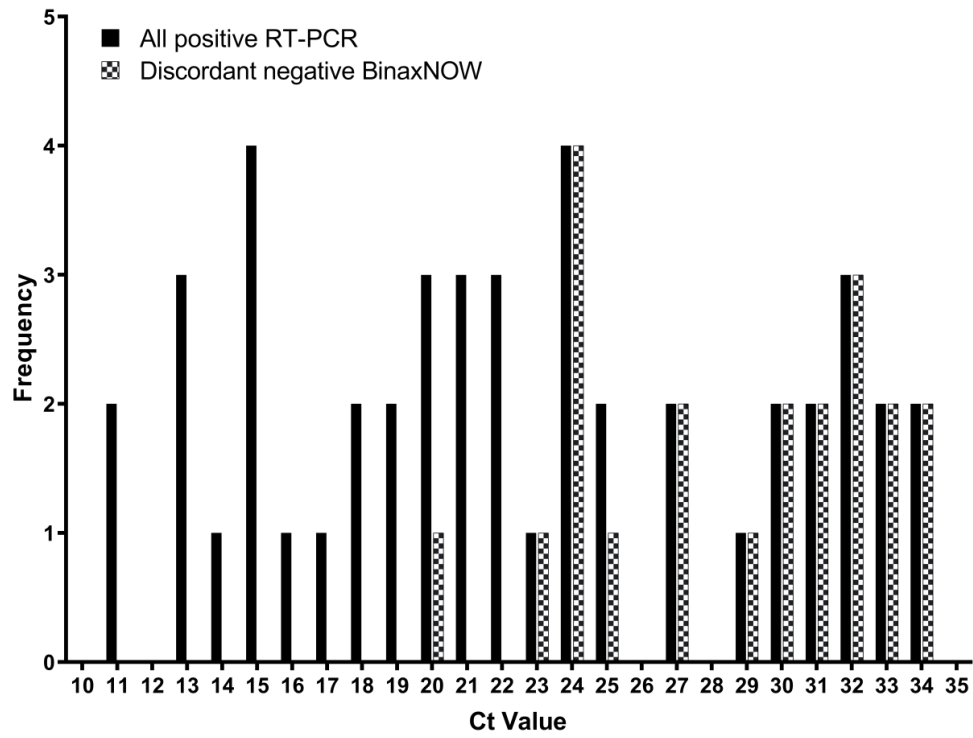
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385 **Figure 2**

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389 **Figure 3**